

John Montilla^{1,2}, Yash Agarwal², Taylor Kist², Peter J. Gaskill², Stephanie M. Matt^{1,2}

Drexel University College of Medicine, ¹Graduate School of Biomedical Sciences and Professional Studies, ²Department of Pharmacology and Physiology

Introduction

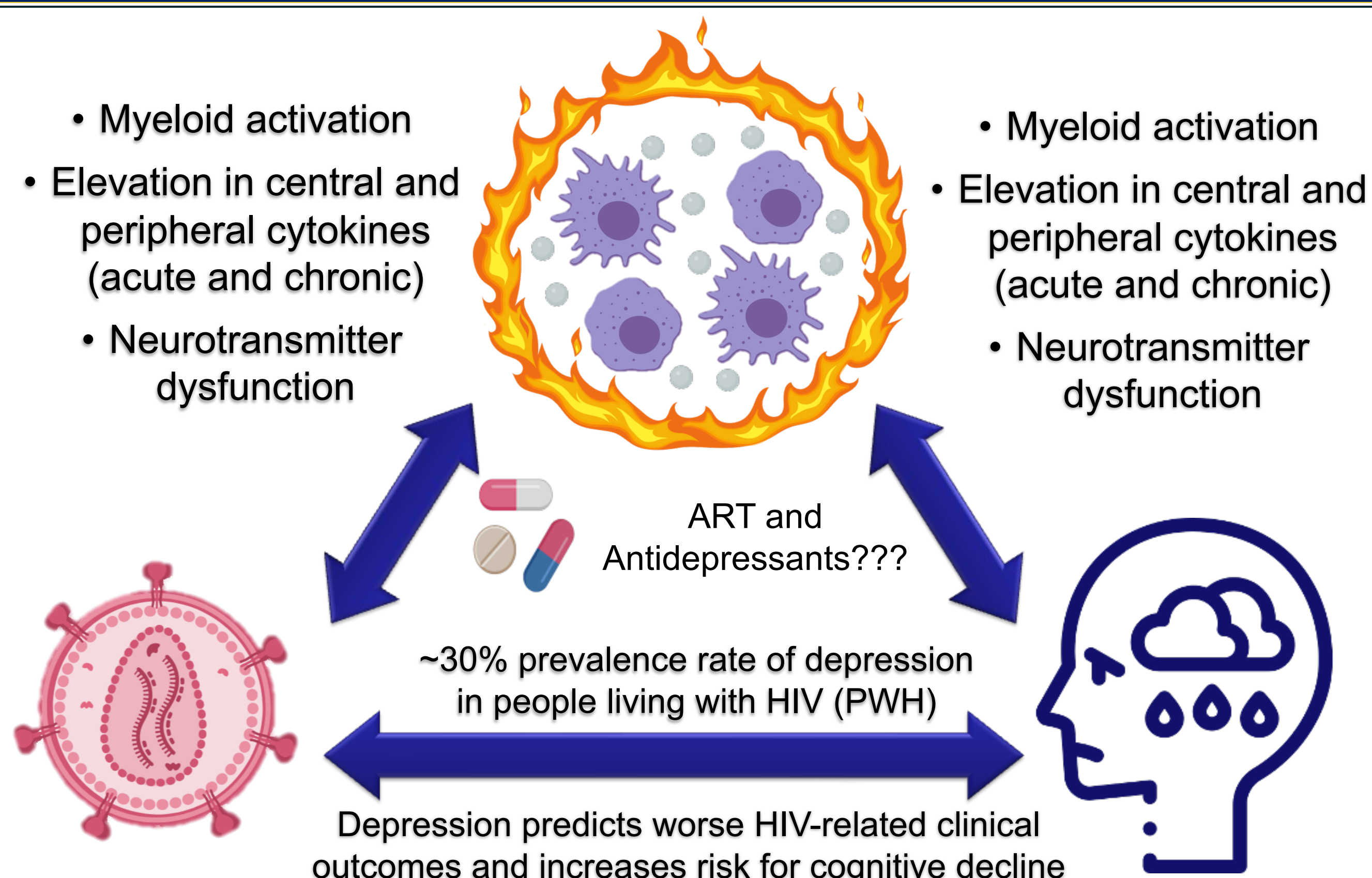
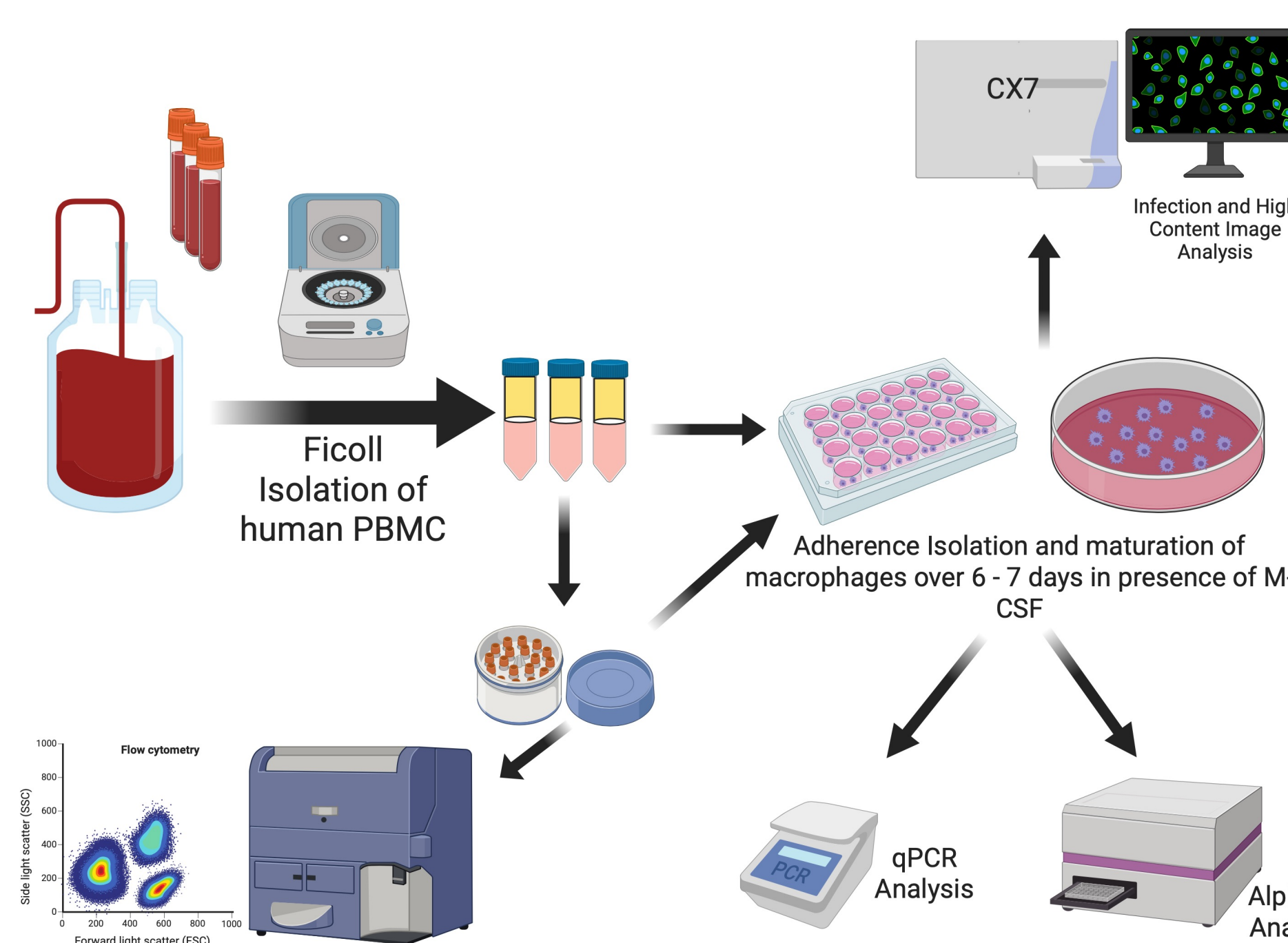


Figure 1: Interactive Impact of Depression, HIV, and Inflammation on Myeloid Cells. Adapted from Rakshasa-Loots et al., 2022. Created with Biorender.

Methods

Figure 2: Experimental Workflow of Processing Fresh and Frozen Monocyte-derived Macrophages. Created with Biorender.



Ficoll-Paque gradient centrifugation separated the peripheral blood mononuclear cells (PBMC) from de-identified blood from the New York Blood Center or virally suppressed HIV+ individuals with/without depression from the Drexel Comprehensive NeuroHIV Center (CNHC). Depression cutoffs were determined PHQ-9. Patient samples were frozen down after isolation and banked through the CNHC until ready to use.

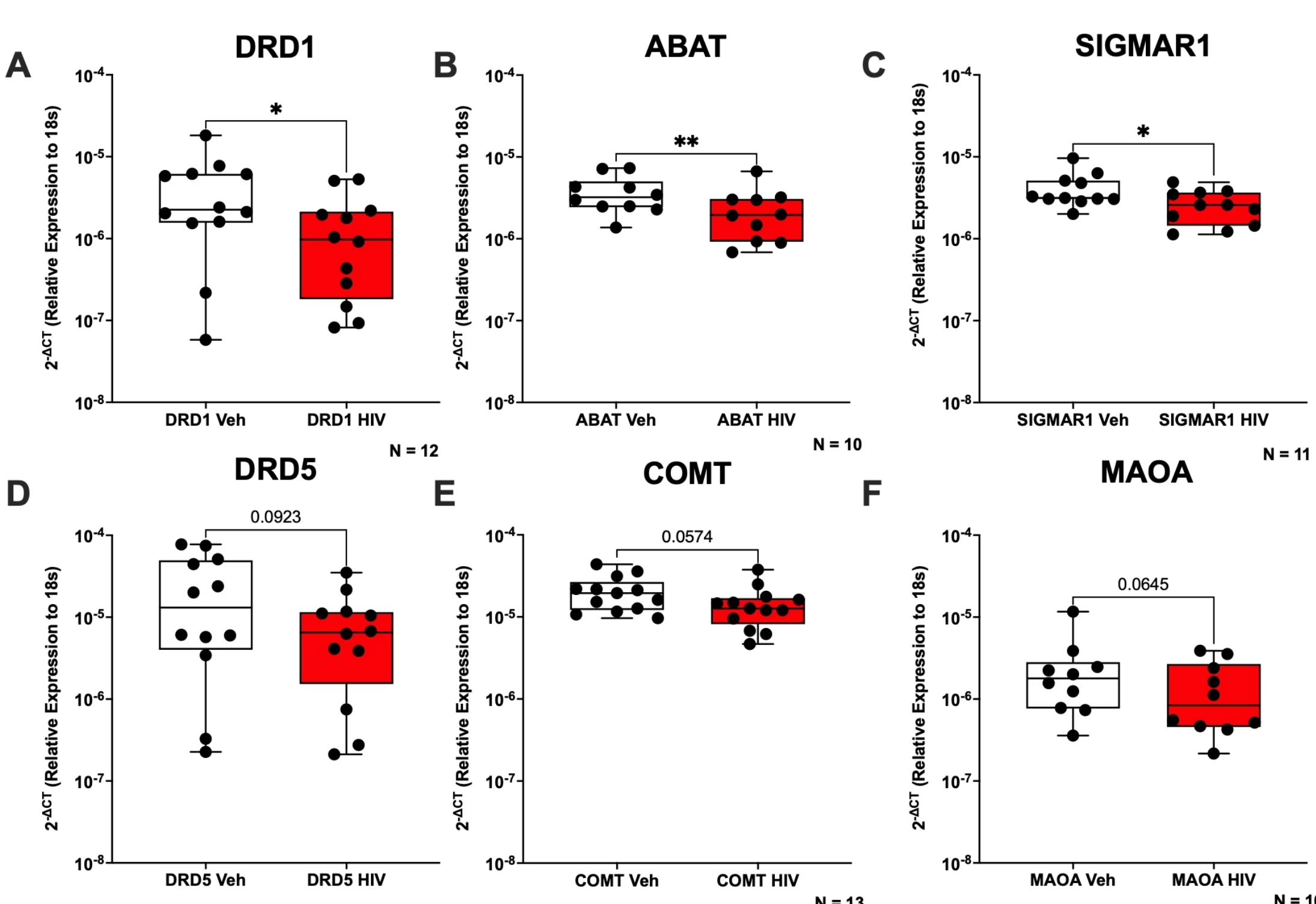
Fresh or thawed PBMC were matured into monocyte-derived macrophages (MDM) using adherence isolation. Cells were cultured for 6-7 days in media containing macrophage colony stimulating factor (M-CSF).

Fresh healthy MDM were mock-infected or infected with 1/ng/mL HIV for 7 days and analyzed for gene expression by qPCR (Figure 3).

PBMC characterization of CNHC samples was analyzed by flow cytometry (Figure 4). Conditions were optimized for frozen MDM (Figure 5) and treated with vehicle, PAMCSK4 (TLR2 Agonist), Poly i:c (TLR3 Agonist), and LPS (TLR4 Agonist) for 1 hour to assess Nf-κB nuclear translocation and 24 hours to assess cytokine release (Figure 6).

Results

Figure 3: Expression of Genes Involved in Neurotransmitter Signaling, Synthesis, and Degradation in HIV-Infected Human Macrophages



Mock-infected or HIV-infected MDM in 6 wells were collected for mRNA analysis. Quantitative RT-PCR detected mRNA for (A) DRD1, (B) ABAT, (C) SIGMAR1, (D) DRD5, (E) COMT, and (F) MAOA. HIV-infected MDM had a significant decrease in DRD1, ABAT, and SIGMAR1. Significance was determined using a Wilcoxon test, *p < 0.05, **p < 0.01.

HIV+ No Depression (N = 11)		HIV+ Depression (N = 19)	
Variable	Statistic	Variable	Statistic
Age (years) ^a	51.5 (6.1) [41-58]	Age (years) ^a	50.1 (7.9) [34-60]
Gender (% men)	50%	Gender (% men)	33.3%
Ethnicity		Ethnicity	
Black/African American	100%	Black/African American	100%
HCV status (% +)	0%	HCV status (% +)	0%
Current CD4 ^a	750.6 (226.1) [495 - 1169]	Current CD4 ^a	799.9 (327.7) [185 - 1332] ^b
ART Status (% on)	100%	ART Status (% on)	100%
PHQ-9 ^a	2.2 (1.5) [0-4]	PHQ-9 ^a	11.3 (5.8) [5-23]

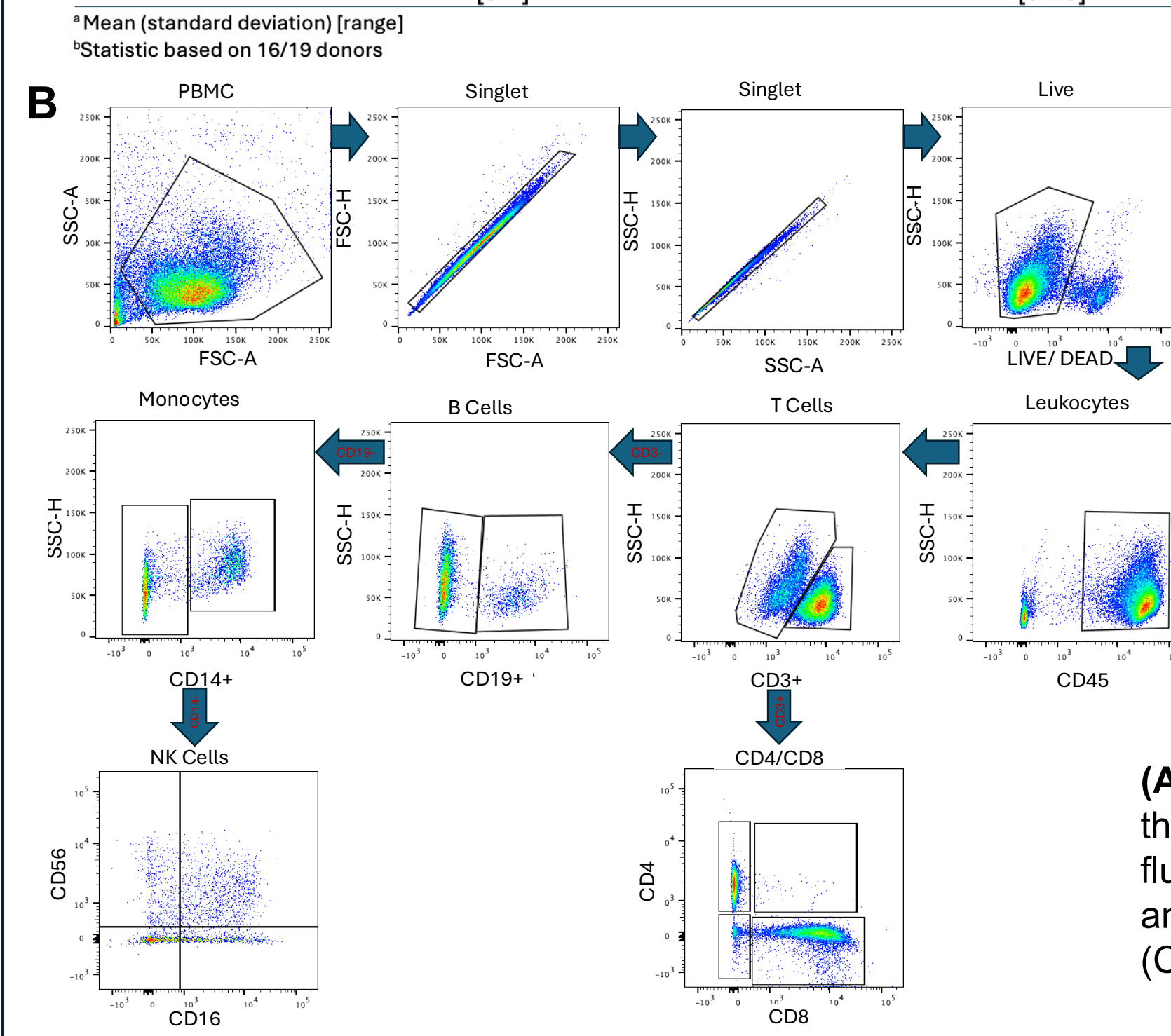
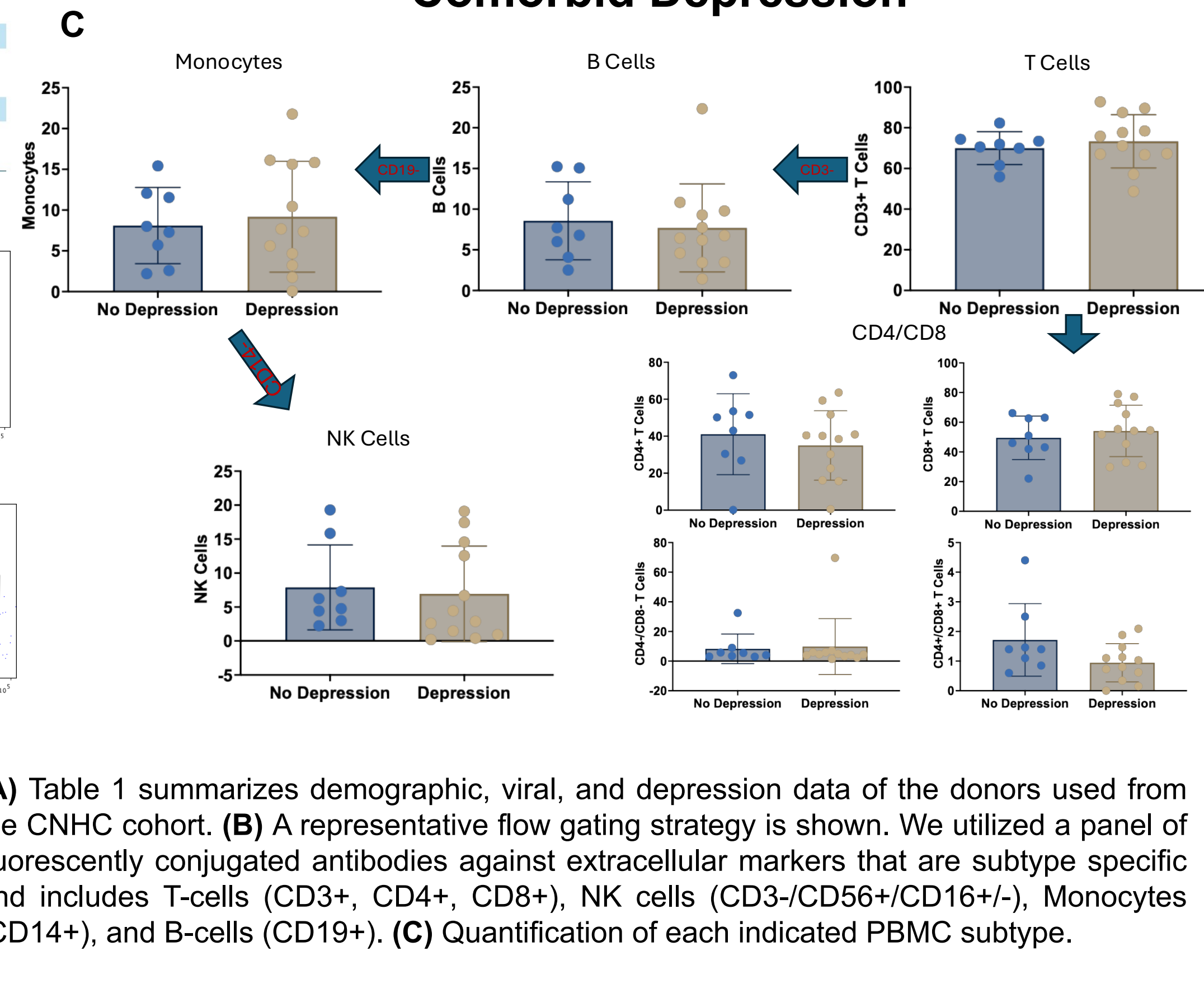
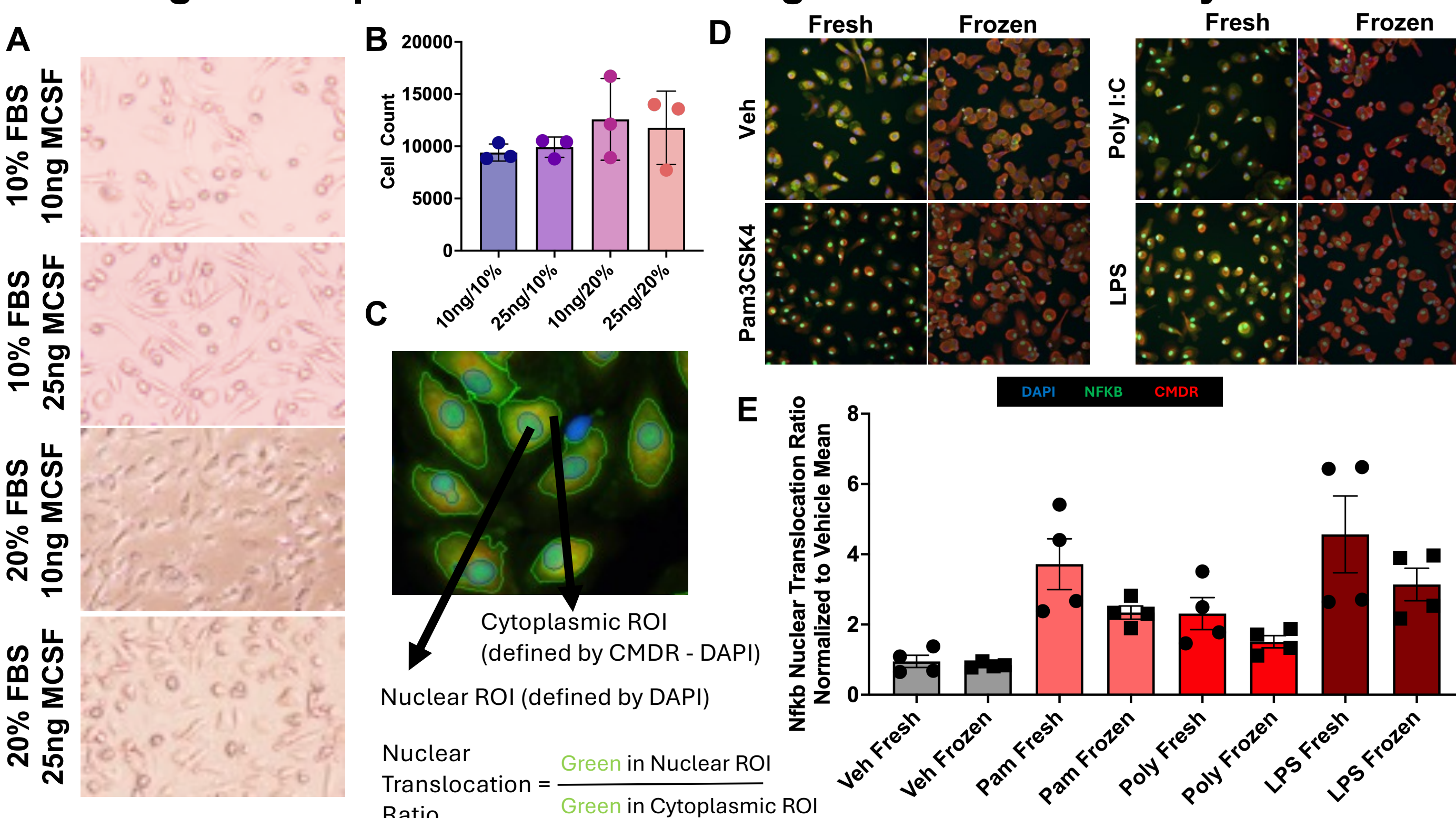


Figure 4: PBMC Characterization of HIV-infected Virally Suppressed Patients with Comorbid Depression



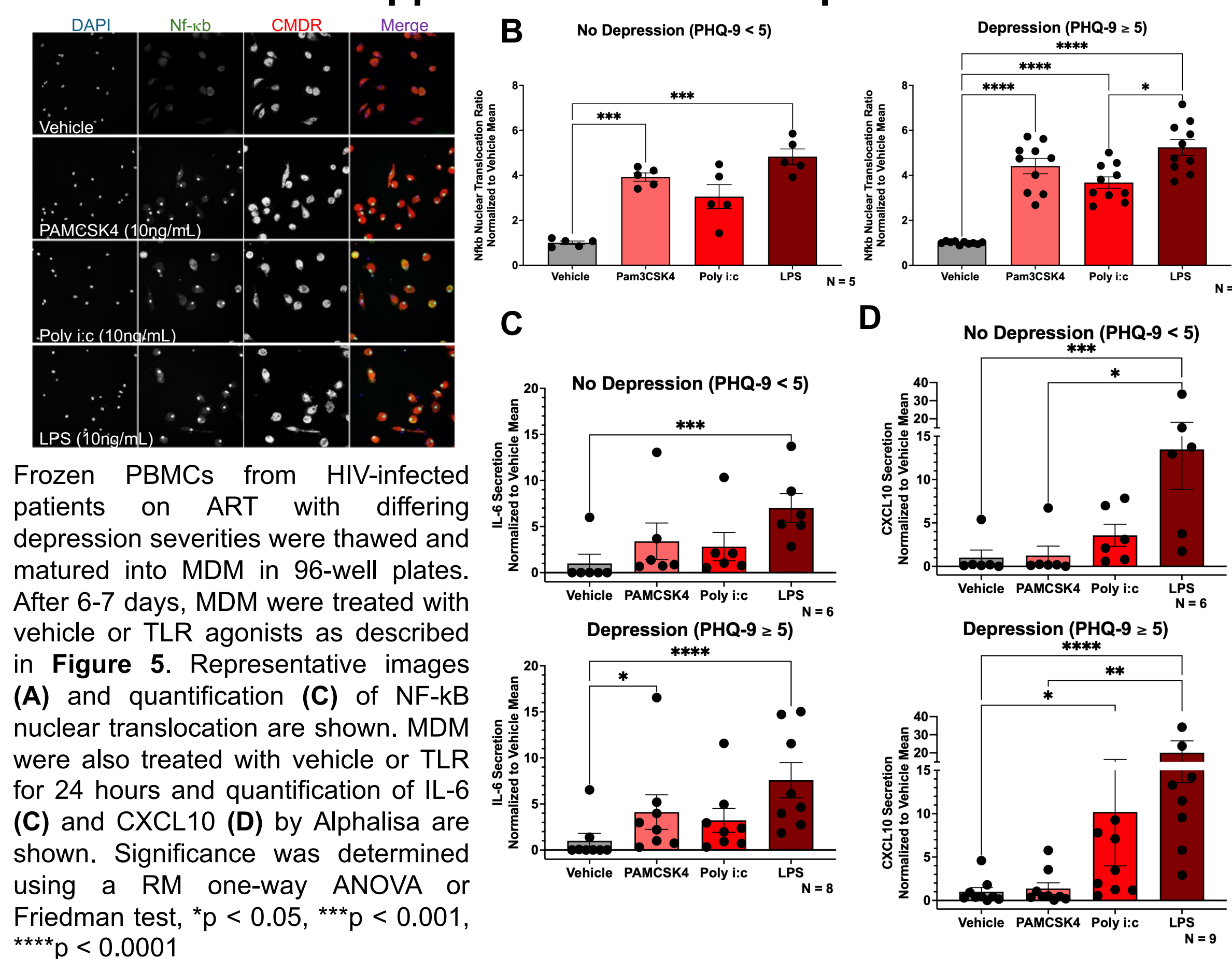
(A) Table 1 summarizes demographic, viral, and depression data of the donors used from the CNHC cohort. (B) A representative flow gating strategy is shown. We utilized a panel of fluorescently conjugated antibodies against extracellular markers that are subtype specific and includes T-cells (CD3+, CD4+, CD8+), NK cells (CD3-/CD56+/CD16+/-), Monocytes (CD14+), and B-cells (CD19+). (C) Quantification of each indicated PBMC subtype.

Figure 5: Optimization of Working with Frozen Monocyte-derived Macrophages



MDM were cultured in 10% or 20% FBS with 10 or 25ng M-CSF in 24-well plates (A). MDM were also either media-fed or washed at days *in vitro* (DIV) 3. Analyses including cell counts (B) determined 10% FBS and 10ng M-CSF with a DIV 3 wash was the optimal condition. Fresh and frozen MDM were treated with vehicle or the TLR agonists PAMCSK4, Poly i:c, or LPS for 1 hour. Description of analysis of Nf-κB nuclear translocation is shown in C. Representative images (D) and quantification (E) of Nf-κB nuclear translocation are shown, with no statistical difference between fresh and frozen responses. Significance was determined using a paired t test, *p < 0.05.

Figure 6: Quantifying Depression-mediated Inflammation by Nf-κB in Macrophages From Frozen HIV-infected Virally Suppressed Patient Samples



Frozen PBMCs from HIV-infected patients on ART with differing depression severities were thawed and matured into MDM in 96-well plates. After 6-7 days, MDM were treated with vehicle or TLR agonists as described in Figure 5. Representative images (A) and quantification (C) of Nf-κB nuclear translocation are shown. MDM were also treated with vehicle or LPS for 24 hours and quantification of IL-6 (C) and CXCL10 (D) by AlphaLisa are shown. Significance was determined using a RM one-way ANOVA or Friedman test, *p < 0.05, ***p < 0.001, ****p < 0.0001

Discussion and Future Directions

- Primary human monocyte-derived macrophages (MDM) express mRNA related to neurotransmitter signaling pathways that significantly change with HIV infection. These studies will guide targets to measure in frozen clinical samples.
- Optimization of frozen MDM demonstrated that 10% FBS and 10ng M-CSF with a DIV 3 wash was the most ideal to move forward analyzing patient-derived samples.
- We can assess PBMC characteristics by flow cytometry as well as Nf-κB nuclear translocation and cytokine release by high-content assays for frozen clinical cohort samples from people living with HIV and depression
- A detailed understanding of depression-mediated changes in inflammation and neurotransmitter signaling will be critical for identifying efficacious therapies to mitigate treatment-resistant depression, suboptimal ART efficacy, and persistent cognitive symptoms in people with HIV.

Acknowledgments

We would like to thank everyone in the Gaskill/Matt labs for their support in this work. We would also like to thank the Comprehensive NeuroHIV Consortium (Drexel University) and its participants for their continued blood donations. Finally, we would like to thank the DUCOM Flow Core and Department of Pharmacology & Physiology for sharing resources, expertise, and equipment. This work was supported by grants from the National Institute on Drug Abuse (R01 DA057337, R61 DA058501), the National Institute of Mental Health (K01 MH132466 to SMM), and the Comprehensive NeuroHIV Center Pilot Grant (SMM).

Dr. Stephanie Matt's Faculty Profile

