FISEVIER

Contents lists available at ScienceDirect

# Schizophrenia Research

journal homepage: www.elsevier.com/locate/schres



# Structural brain alterations in schizophrenia following fetal exposure to the inflammatory cytokine interleukin-8

Lauren M. Ellman <sup>a,\*</sup>, Raymond F. Deicken <sup>b</sup>, Sophia Vinogradov <sup>b,c</sup>, William S. Kremen <sup>d</sup>, John H. Poole <sup>e</sup>, David M. Kern <sup>h</sup>, Wei Yann Tsai <sup>f</sup>, Catherine A. Schaefer <sup>g</sup>, Alan S. Brown <sup>h,i</sup>

- <sup>a</sup> Department of Psychology, Temple University, Philadelphia, PA, United States
- <sup>b</sup> Department of Psychiatry, University of California San Francisco, San Francisco, CA, United States
- <sup>c</sup> Mental Health Service, San Francisco Department of Veteran Affairs Medical Center, San Francisco, CA, United States
- <sup>d</sup> Department of Psychiatry, Center for Behavioral Genomics, University of California San Diego, and Center of Excellence for Stress and Mental Health, VA San Diego Healthcare System, La Jolla, CA, United States
- Department of Neuropsychology, Defense & Veterans Brain Injury Center, Veterans Affairs Health Care System, Palo Alto, CA, United States
- f Department of Biostatistics, Mailman School of Public Health, Columbia University, United States
- <sup>g</sup> Kaiser Permanente Division of Research, Oakland, CA, United States
- h Department of Psychiatry, College of Physicians and Surgeons of Columbia University, New York State Psychiatric Institute, New York, NY, United States
- <sup>i</sup> Department of Epidemiology, Mailman School of Public Health, Columbia University, New York, NY, United States

#### ARTICLE INFO

Article history: Received 27 October 2009 Received in revised form 11 May 2010 Accepted 12 May 2010 Available online 9 June 2010

Keywords:
Obstetric complications
Schizophrenia
Proinflammatory cytokines
Structural MRI
Pregnancy
Infection

#### ABSTRACT

Background: Maternal infection during pregnancy has been repeatedly associated with increased risk for schizophrenia. Nevertheless, most viruses do not cross the placenta; therefore, the damaging effects to the fetus appear to be related to maternal antiviral responses to infection (e.g. proinflammatory cytokines). Fetal exposure to the proinflammatory cytokine interleukin-8 (IL-8) has been significantly associated with risk of schizophrenia in offspring. This study sought to determine the association between fetal exposure to IL-8 and structural brain changes among schizophrenia cases and controls.

Methods: Subjects were 17 cases diagnosed with schizophrenia from the Developmental Insult and Brain Anomaly in Schizophrenia (DIBS) study. Psychiatric diagnoses were determined among offspring with semi-structured interviews and medical records review. IL-8 was determined from assays in archived prenatal sera and volumetric analyses of neuroanatomical regions were obtained from T1-weighted magnetic resonance imaging in adulthood. Eight controls were included for exploratory purposes.

Results: Among cases, fetal exposure to increases in IL-8 was associated with significant increases in ventricular cerebrospinal fluid, significant decreases in left entorhinal cortex volumes and significant decreases in right posterior cingulate volumes. Decreases that approached significance also were found in volumes of the right caudate, the putamen (bilaterally), and the right superior temporal gyrus. No significant associations were observed among controls.

Conclusion: Fetal exposure to elevations in maternal IL-8 led to structural neuroanatomic alterations among cases in regions of the brain consistently implicated in schizophrenia research. In utero exposure to elevations in IL-8 may partially account for brain disturbances commonly found in schizophrenia.

© 2010 Elsevier B.V. All rights reserved.

E-mail address: ellman@temple.edu (L.M. Ellman).

#### 1. Introduction

Previous studies have found structural changes throughout the brain in schizophrenia patients compared to non-

<sup>\*</sup> Corresponding author. Temple University, Weiss Hall, 1701 North 13th Street, Philadelphia, PA 19122-6085, United States. Tel.: +1 215 204 1571; fax: +1 215 204 5539.

psychiatric controls. Among these studies, increased ventricular size is the most well-replicated structural anomaly found in schizophrenia research (Gur et al., 2007; Wright et al., 2000). Decreases in whole brain volumes and temporolimbic regions also have been repeatedly found among schizophrenia patients (Gur et al., 2007; Wright et al., 2000). Further, studies of individuals who are in the prodrome of schizophrenia also have identified multiple structural brain changes, suggesting that brain disturbances associated with the disorder may have neurodevelopmental origins (Job et al., 2005; Pantelis et al., 2003). Despite these findings, few investigations have sought to determine the contributions of environmental risk factors to neuroanatomical changes found in schizophrenia.

Although genetic factors are believed to substantially contribute to the etiology of schizophrenia, concordance rates approximating 50% between monozygotic twins indicates the presence of substantial environmental influences (Cannon et al., 1998). Among the possible environmental contributors, maternal infections during pregnancy have been repeatedly linked to an increased risk for schizophrenia (Brown and Derkits, 2010). Nevertheless, many infections do not appear to cross the placenta; therefore the damaging influences to the fetal brain seem related to maternal antiviral responses to infection, such as increases in proinflammatory cytokines (Patterson, 2009). In a previous study using the birth cohort of the current investigation, increases in maternal levels of interleukin-8 (IL-8) during the second/third trimesters of pregnancy were associated with increased risk for schizophrenia among offspring (Brown et al., 2004).

Hence, we sought to examine whether fetal exposure to increases in maternal IL-8 during the second/third trimesters results in more pronounced structural brain alterations among individuals diagnosed with schizophrenia and other spectrum disorders (herein referred to as schizophrenia). Our primary hypothesis was that fetal exposure to IL-8 would result in increases in ventricular cerebrospinal fluid (CSF) volume among cases. In addition to the well-replicated association between increases in ventricular CSF and schizophrenia, this hypothesis was derived from animal studies indicating increased ventricular volumes following fetal exposure to immune activation (Patterson, 2009; Wright et al., 2000). Based on previous findings from studies of schizophrenia patients, prodromal research and animal studies, we also predicted that fetal exposure to increased maternal IL-8 would be associated with reduced volumes of temporal lobe regions, particularly in the hippocampus, parahippocampus, and the superior temporal gyrus, and reductions in basal ganglia volumes (Pantelis et al., 2003; Patterson, 2009; Wright et al., 2000). Exploratory analyses were conducted with control participants and on additional regions of interest (ROIs). Analyses of controls were conducted to obtain preliminary findings on whether there are differential vulnerabilities to fetal exposure to IL-8 among cases versus controls, as has been found in other studies (Cannon et al., 2008; Ellman et al., 2009).

## 2. Materials and methods

All subjects provided written informed consent and the study was approved by the Institutional Review Boards of the New York State Psychiatric Institute, the Kaiser Foundation Research Institute, and the University of California San Francisco VA Medical Center.

#### 2.1. Description of the cohort

The subjects were derived from the Developmental Insult and Brain Anomaly in Schizophrenia (DIBS) study that was based on participants from the Prenatal Determinants of Schizophrenia (PDS) study, which ascertained cases of schizophrenia from a large birth cohort described in detail previously (Susser et al., 2000). Briefly, the PDS study included pregnant women ( $n\!=\!12,094$  live births) receiving obstetric care from the Kaiser Permanente Medical Care Plan (KPMCP) in Alameda County, California, as part of the Child Health and Development Study (CHDS). Maternal serum samples were collected during the pregnancies and were frozen and stored at  $-20\,^{\circ}$  C.

#### 2.2. Ascertainment and diagnosis

Case ascertainment and screening were accomplished following computerized record linkage between the CHDS and KPMCP identifiers from inpatient, outpatient, and pharmacy registries of CHDS participants who belonged to KPMCP from 1981 to 1997 (corresponding to the initiation of KPMCP computerized psychiatric registries and to the end of follow-up). Potential cases were diagnosed using DSM-IV criteria following assessment with the Diagnostic Interview for Genetic Studies (DIGS) (Nurnberger et al., 1994), chart review, and consensus of 3 experienced research psychiatrists.

All cases and controls were targeted for neuroimaging assessments in adulthood. DIBS study participants consisted of 23 cases of schizophrenia and 25 controls for MRI acquisition and analysis. Two cases were excluded due to unusable images from movement artifacts. The final sample was comprised of 17 cases and 8 controls with available MRI and available second/ third trimester cytokine data. Among the 17 cases, 7 were diagnosed with schizophrenia, 4 with schizoaffective disorder, and 6 with other schizophrenia spectrum disorders. Demographic characteristics of the cases and controls are provided in Table 1. There were no significant differences in any demographic characteristics between cases and controls. There also were no significant differences in any demographic variables between DIBS cases and cases from the PDS study with available cytokine data that were not ascertained as part of the DIBS study (Table 2).

#### 2.3. Interleukin-8 assay

Analyses were restricted to second/third trimester IL-8 values, because 1) maternal IL-8 during the second/third trimester was previously associated with schizophrenia in this birth cohort, whereas other proinflammatory cytokines (IL-6, TNF- $\alpha$ , IL-1 $\beta$ ) were not associated with schizophrenia (Brown et al., 2004) and 2) we sought to reduce the probability of Type I errors by limiting the number of analyses.

Assays were conducted blind to diagnosis of the offspring and were carried out under Level II biohazard containment conditions. Each serum specimen was thawed and made 2 mM with respect to phenylmethylsulfonyl fluoride, a protease inhibitor, to prevent degradation of cytokines by serum

**Table 1** Demographic characteristics of sample.

Characteristic	SSD $(n=17)$	No diagnosis $(n=8)$	<i>p</i> -value	
% Female	5 (29.41%)	2 (25%)	0.819	
Age at MRI scan	39.96 (1.78)	41.17 (1.69)	0.140	
Birth weight	3508.73 (522.13)	3378.38 (572.20)	0.323	
Gestational length (weeks)	40.39 (1.58)	40.14 (0.71)	0.722	
Maternal age	28.12 (6.54)	26.88 (6.58)	0.662	
Maternal earlier deliveries	1.88 (2.18)	0.75 (0.89)	0.173	
Maternal education <=HS graduate*	12 (70.59)	4 (66.67)	0.858	
Maternal race*				
White	12 (70.59%)	7(87.50%)	0.096	
Black	5 (29.41%)	0		
Asian	0	1 (12.50%)		
IL-8 (pg/ml)	952.04 (1083.45)	315.24 (248.60)	0.118	
Medication use				
Atypical antipsychotics	4 (23.53)	0		
Typical antipsychotics	3 (17.65)	0		
On any antipsychotic	7 (18.92)	0		
Anti-pyramidal drugs	3 (17.65)	0		
Mood stabilizers	2 (11.76)	0		
SSRIs	2 (11.76)	0		
Other anti-depressant (trazadone)	1 (5.88)	0		
Ritalin	1 (5.88)	0		
Methadone	1 (5.88)	0		

<sup>\*</sup>Information was incomplete for 2 DIBS controls on maternal education and for 3 DIBS gravidas maternal race was incomplete; therefore this variable was extrapolated from child's race for these gravidas.

proteases. Measurement of IL-8 was conducted using the sandwich enzyme-linked immunosorbent assay described in detail in a previous study from this cohort (Brown et al., 2004).

#### 2.4. Image acquisition and analysis

MR images were acquired using a 1.5-Tesla Siemens system. Coronal T1-weighted images were obtained from 3D MP-RAGE sequences (TR/TI/TE = 10/250/4 ms, resolution  $1\times1$  mm², 1.4 mm slice thickness). MRI tissue segmentation and regional voluming in the Talairach coordinate system were used based on the cited methods (Collins et al., 1998; Kwan et al., 1999). These methods have been shown to be reliable (Manji et al., 2000) and valid (Fein et al., 2000) in previous studies. In-house software was used to: 1) remove the skull and meninges from the images; 2) coregister each of the interleaves of the spin-

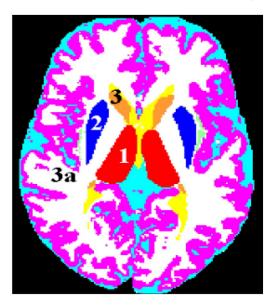
echo images to T1 images reformatted to the axial plane; 3) perform RF inhomogeneity correction in 3D; 4) define seeds (based on peaks in the 3D histogram of T1 pixel intensities) for the K-means cluster analysis; and 5) transfer the data to statistical software which performs the actual cluster analysis. The initial process was followed by computer-assisted editing of the data to separate cortical from subcortical gray matter, ventricular CSF from sulcal CSF, and to reclassify white matter incorrectly classified in the first pass into a category of white matter with an abnormal MRI signal or white matter signal hyperintensity (WMSH). This was followed by manual delineation of the boundaries of cortical regions, subcortical structures, the cerebellum, and the hippocampi. The transformation to the Talairach coordinate system involved piecewise linear transformations of 12 compartments for each subject's brain. Each subject's tissue contribution to the commonly

**Table 2**Demographic characteristics of DIBS and non-DIBS cases.

Characteristic	DIBS cases $(n=17)$	Non-DIBS cases ** (n = 42)	<i>p</i> -value
% Female	5 (29.41%)	15 (35.71%)	0.643
Birth weight (g)	3508.73 (522.13)	3332.48(562.67)	0.245
Gestational length (weeks)*	40.39 (1.58)	40.59 (2.26)	0.738
Maternal age	28.12 (6.54)	27.91 (5.59)	0.90
Maternal earlier deliveries	1.88 (2.18)	2.05 (1.90)	0.77
Maternal education <=HS graduate*	12 (70.59)	24 (66.67)	0.775
Maternal race*			
White	12 (70.59%)	17 (41.46%)	0.775
Black	5 (29.41%)	23 (56.10%)	
Asian	0	1 (2.443%)	
IL-8 (pg/ml)	952.04 (1083.45)	1020.81(1644.09)	0.875

<sup>\*</sup>Information was incomplete for 8 non-DIBS cases on maternal education, for 1 non-DIBS case on gestational length, and for 1 non-DIBS case on maternal and child race. For 3 DIBS gravidas maternal race was incomplete; therefore this variable was extrapolated from child's race for these gravidas.

<sup>\*\*</sup>Non-DIBS cases were defined as any cases of schizophrenia with available IL-8 data who were part of the larger PDS cohort, but not ascertained in the present t2:16 study.



**Fig. 1.** Axial MRI tissue segmented image. This figure displays manually segmented areas of the thalamus (1); putamen (2); head of caudate (3); tail of caudate (3a).

defined ROI was then computed by superimposing the subject-specific ROI on the subject's segmented image, and counting the (segmented) pixels contained in the ROI. Using this approach, reliability studies for 20 normal subjects on two separate occasions revealed inter- and intraoperator correlations of 0.91 to 0.99 for ventricular CSF, sulcal CSF, cortical gray matter, white matter, and % white matter signal hyperintensities. All cortical volumes, including superior temporal gyrus (Brodmann area 22), entorhinal cortex (Brodmann area 28), inferior frontal cortex (Brodmann areas 44 and 45), dorsolateral prefrontal cortex (DLPFC, Brodmann areas 9 and 46), anterior cingulate (Brodmann areas 24, 25, and 32), and posterior cingulate (Brodmann area 31) were obtained based on the methods above.

The thalamus (see Fig. 1; Deicken et al., 2002), and hippocampus (see Fig. 2; Deicken et al., 1999; Talairach and Tournoux, 1988) were manually traced using the cited



**Fig. 2.** ROI tracing of hippocampus. This figure shows a representative coronal MR image with the hippocampal areas shown in yellow.

methods. Reliability studies of hippocampal and thalamic volumes (n = 10) of normal controls revealed interoperator correlations of 0.92 to 0.96.

The head of the caudate (see Fig. 1) was defined on the transaxial plane as the mass of gray matter comprising the lateral walls of the lateral ventricles bounded inferiorly and laterally by the anterior limb of the internal capsule and superiorly and laterally by the external capsule. Region placement for the caudate and the putamen began inferiorly when the operator could see clearly at least on one side the anterior limb of the internal capsule dividing the caudate head and the putamen. The boundary for the head of the caudate continued superiorly until the thalamus could no longer be visualized and the anterior horn of the lateral ventricles became confluent with the posterior horn. The body of the caudate was defined as the portion above the thalamus after the confluence of the anterior and posterior horns of the lateral ventricles. The caudate tail was defined as a structure of gray matter closely adjacent anterolaterally to the posterior horns of the lateral ventricles and posterolaterally to the thalamus until it became confluent with the body of the caudate. Reliability studies of caudate volumes (n=10) of normal controls revealed interoperator correlations of 0.92 to 0.93.

The putamen (see Fig. 1) is bounded laterally by the external capsule, and postero- and antero-medially by the internal capsule. The boundary between the putamen and the globus pallidus is noted by the difference between the two structures and when possible a strip of white matter was identified between the gray matter masses of the two structures. Regional tracing for the putamen extended superiorly until no more gray matter pixels could be detected medially to the claustrum band. Reliability studies of putamen volumes ( $n\!=\!10$ ) of normal controls revealed interoperator correlations of 0.93 to 0.94.

All ROIs were divided by intracranial volume to correct for head size. Ratios were chosen instead of controlling for intracranial volumes, in order to minimize degrees of freedom given the modest sample size.

### 2.5. Statistical analyses

As is often the case with cytokine distributions, there was a wide range of IL-8 values (3.63 pg/ml to 3717.7 pg/ml) and this distribution was skewed. IL-8 values were log-transformed due to this non-normal distribution, as well as ease of interpretation. Multiple regressions were conducted in SAS version 9.1 (SAS, Inc., Cary, N.C.). Standardized parameter estimates were calculated in addition to traditional unstandardized estimates to allow for interpretation of the magnitude of the relationship between IL-8 and volumetric measures, as represented in standardized units. Standardized estimates represent the standard deviation changes in volumetric ratios that result from a change of one standard deviation in log IL-8 values. Maternal education at birth (HS graduate or below versus higher education) was added to the models to control for the potential influences of postnatal adversity. Maternal education has been highly correlated with other measures of socioeconomic status in the PDS study and is often used as a proxy variable for postnatal adversity in studies of prenatal risk factors to control for the potential contributions of postnatal disadvantages to brain alterations (Schlotz and Phillips, 2009). Age of participants, participants' sex, and medication status were explored as potential control variables and/or confounders by examining the relationship between these variables and IL-8 levels and volumetric measures. Statistical significance was based on p<0.05; all tests were two-tailed. Primary analyses included all analyses with a priori, directional hypotheses; whereas secondary analyses were used for exploratory purposes.

#### 3. Results

There were no significant differences in age or sex between cases and controls (see Table 1). In addition, among several

demographic variables examined, age (r=0.0575, p=0.785), (F=0.35, df=1, 24, p=0.559), and medication status of cases (on antipsychotics or not; F=0.01, df=1, 24, p=0.924) were not significantly related to IL-8 levels. Further, there were no differences in brain volumes between cases that were and were not taking antipsychotics at the time of the assessment (results available upon request).

In primary analyses among cases (see Table 3 and Fig. 3), fetal exposure to increases in maternal IL-8 levels was significantly associated with increases in ventricular CSF volume among cases (p = 0.045). Fetal exposure to IL-8 also was related to significant decreases in left entorhinal cortex volumes (p = 0.037) and decreases that approached signifi-

**Table 3**Results of unadjusted and adjusted regression analyses of maternal IL-8 and brain volumes in cases and controls: primary analyses.

Brain region	Control participants (n=8) unadjusted models				Control participants ( $n = 8$ ) models adjusted for maternal education				
	Parameter estimate	95% CI	β	p-value	Parameter estimate	95% CI	β	p-value	
Ventricular regions									
Ventricular cerebrospinal fluid	0.002	- 0.006, 0.011	0.249	0.552	-0.003	-0.023, 0.016	- 0.386	0.631	
Cortical/limbic structures									
Entorhinal cortex (right)	0.000005	-0.0003, 0.0003		0.975	0.0002	-0.0004, 0.0008	0.731	0.393	
Entorhinal cortex (left)	-0.00004	-0.0004, 0.0003	-0.097	0.820	0.0002	-0.0007, 0.001	0.481	0.562	
Hippocampus (right)	0.0002	-0.00003, 0.0004	0.649	0.082†	0.0004	-0.00003, 0.0008	1.281	0.061†	
Hippocampus (left)	-0.00001	-0.0002, 0.0001	-0.077	0.856	-0.00002	-0.0004, 0.0004	-0.166	0.858	
Superior temporal gyrus (right)	0.000002	-0.001, 0.001	0.002	0.996	0.0005	-0.002, 0.002	0.544	0.518	
Superior temporal gyrus (left)	-0.0001	-0.001, 0.001	-0.104	0.806	0.0004	-0.002, 0.003	0.377	0.661	
Subcortical structures									
Caudate (right)	0.0001	-0.0005, 0.0007	0.184	0.662	0.0002	-0.0001, 0.0006	0.978	0.142	
Caudate (left)	0.0002	-0.0002, 0.0005	0.449	0.264	0.0002	-0.0005, 0.0008	0.471	0.508	
Putamen (right)	0.0001	-0.0004, 0.0006	0.238	0.571	0.0002	-0.0009, 0.001	0.508	0.573	
Putamen (left)	0.0002	-0.0003, 0.0006	0.367	0.371	0.0003	-0.0007, 0.001	0.759	0.381	
					· · · · · · · · · · · · · · · · · · ·				
Brain region	Schizophrenia c	ases ( $n = 17$ ) unadjusted me	odels		Schizophrenia o	cases ( $n = 17$ ) models adjust	ed for mat	ernal	
Brain region	Schizophrenia c Parameter estimate	ases (n=17) unadjusted mo	ρdels	<i>p</i> -value	•	eases (n = 17) models adjust	ed for mat $\beta$		
	Parameter			<i>p</i> -value	education Parameter			p-value	
Ventricular regions	Parameter		β	<i>p</i> -value 0.055†	education Parameter		β		
Ventricular regions Ventricular cerebrospinal fluid	Parameter estimate	95% CI	β		education Parameter estimate	95% CI	β	p-value	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures	Parameter estimate	95% CI	β	0.055†	education Parameter estimate	95% CI	β	<i>p</i> -value 0.045*	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right)	Parameter estimate  0.002	95% CI - 0.00005, 0.004 - 0.00007, 0.00004 - 0.00009,	β 0.473	0.055†	Parameter estimate 0.002	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009,	β 0.487	p-value 0.045*	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left)	Parameter estimate  0.002  - 0.00001 - 0.00005	95% CI - 0.00005, 0.004 - 0.00007, 0.00004 - 0.00009, - 0.000005	β 0.473 -0.138 -0.525	0.055† 0.597 0.030*	education  Parameter estimate  0.002  - 0.00001 - 0.00005	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009, - 0.000003	β 0.487 - 0.124 - 0.519	p-value 0.045* 0.630 0.037*	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right)	Parameter estimate  0.002  -0.00001 -0.00005 0.00002	95% CI  -0.00005, 0.004  -0.00007, 0.00004 -0.00009, -0.000005 -0.00005, 0.00009	$\beta$ 0.473 -0.138 -0.525 0.172	0.055† 0.597 0.030* 0.509	education Parameter estimate  0.002  - 0.00001 - 0.00005 0.00002	95% CI  0.00005, 0.004  -0.00007,0.00004 -0.00009, -0.000003 -0.00005, 0.0001	β 0.487 -0.124 -0.519 0.169	p-value 0.045* 0.630 0.037* 0.531	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus	Parameter estimate  0.002  - 0.00001 - 0.00005	95% CI - 0.00005, 0.004 - 0.00007, 0.00004 - 0.00009, - 0.000005	β 0.473 -0.138 -0.525	0.055†  0.597 0.030*  0.509 0.588	education  Parameter estimate  0.002  - 0.00001 - 0.00005	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009, - 0.000003	β 0.487 - 0.124 - 0.519	p-value 0.045* 0.630 0.037* 0.531 0.523	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right)	Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00002 -0.00002	95% CI  - 0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.00005, 0.00009 - 0.0001, 0.00006 - 0.0004, 0.00007	$\begin{array}{c} \beta \\ \\ -0.138 \\ -0.525 \\ \\ 0.172 \\ -0.142 \\ -0.353 \end{array}$	0.055†  0.597 0.030*  0.509 0.588 0.165	education  Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00003 -0.00002	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009, - 0.000003 - 0.00005, 0.0001 - 0.0001, 0.00006 - 0.0004, 0.00002	β 0.487 -0.124 -0.519 0.169 -0.160 -0.379	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079†	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right)	Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00002	95% CI  - 0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.00005, 0.00009 - 0.0001, 0.00006	$\begin{array}{c} \beta \\ \\ 0.473 \\ \\ -0.138 \\ -0.525 \\ \\ 0.172 \\ -0.142 \end{array}$	0.055†  0.597 0.030*  0.509 0.588 0.165	education Parameter estimate  0.002  - 0.00001 - 0.00005  0.00002 - 0.00003	95% CI  0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.000003 - 0.00005, 0.0001 - 0.0001, 0.00006	β 0.487 - 0.124 - 0.519 0.169 - 0.160	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079†	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right) Superior temporal gyrus (left)	Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00002 -0.00002	95% CI  - 0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.00005, 0.00009 - 0.0001, 0.00006 - 0.0004, 0.00007	$\begin{array}{c} \beta \\ \\ -0.138 \\ -0.525 \\ \\ 0.172 \\ -0.142 \\ -0.353 \end{array}$	0.055†  0.597 0.030*  0.509 0.588 0.165	education  Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00003 -0.00002	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009, - 0.000003 - 0.00005, 0.0001 - 0.0001, 0.00006 - 0.0004, 0.00002	β 0.487 -0.124 -0.519 0.169 -0.160 -0.379	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079†	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right) Superior temporal gyrus (left) Subcortical structures	Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00002 -0.00002	95% CI  - 0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.00005, 0.00009 - 0.0001, 0.00006 - 0.0004, 0.00007 - 0.0003, 0.0001	$\begin{array}{c} \beta \\ \\ -0.138 \\ -0.525 \\ \\ 0.172 \\ -0.142 \\ -0.353 \end{array}$	0.055†  0.597 0.030*  0.509 0.588 0.165  0.390	education  Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00003 -0.00002	95% CI  0.00005, 0.004  -0.00007,0.00004 -0.00009, -0.00003 -0.00005, 0.0001 -0.0001, 0.00006 -0.0004, 0.00002 -0.0003, 0.0001	β 0.487 -0.124 -0.519 0.169 -0.160 -0.379	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079† 0.320	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right) Superior temporal gyrus (left) Subcortical structures Caudate (right)	Parameter estimate  0.002  - 0.00001 - 0.00005  0.00002 - 0.00002 - 0.00002 - 0.00002	95% CI  - 0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.00005, 0.00009 - 0.0001, 0.00006 - 0.0004, 0.00007	β 0.473 - 0.138 - 0.525 0.172 - 0.142 - 0.353 - 0.223	0.055†  0.597 0.030*  0.509 0.588 0.165 0.390  0.042*	education  Parameter estimate  0.002  - 0.00001 - 0.00005  0.00002 - 0.00003 - 0.0002 - 0.0001	95% CI  0.00005, 0.004  - 0.00007,0.00004 - 0.00009, - 0.000003 - 0.00005, 0.0001 - 0.0001, 0.00006 - 0.0004, 0.00002	β 0.487 -0.124 -0.519 0.169 -0.160 -0.379 -0.242	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079† 0.320	
Ventricular regions Ventricular cerebrospinal fluid Cortical/limbic structures Entorhinal cortex (right) Entorhinal cortex (left) Hippocampus (right) Hippocampus (left) Superior temporal gyrus (right) Superior temporal gyrus	Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00002 -0.0002 -0.0001  -0.00006	95% CI  -0.00005, 0.004  -0.00007, 0.00004 -0.00009, -0.00005, 0.00009 -0.0001, 0.00006 -0.0004, 0.00007 -0.0003, 0.0001	$\beta \\ 0.473 \\ -0.138 \\ -0.525 \\ 0.172 \\ -0.142 \\ -0.353 \\ -0.223 \\ -0.497$	0.055†  0.597 0.030*  0.509 0.588 0.165 0.390  0.042* 0.573	education  Parameter estimate  0.002  -0.00001 -0.00005  0.00002 -0.00003 -0.0002 -0.00001	95% CI  0.00005, 0.004  - 0.00007, 0.00004 - 0.00009, - 0.000003 - 0.00005, 0.0001 - 0.0001, 0.00006 - 0.0004, 0.00002  - 0.0003, 0.0001	β 0.487 -0.124 -0.519 0.169 -0.160 -0.379 -0.242 -0.495	p-value 0.045* 0.630 0.037* 0.531 0.523 0.079† 0.320 0.051† 0.594	

This table includes the results from the primary multiple regression analyses with and without controlling for maternal education at birth. Unstandardized parameter estimates, 95% confidence intervals of the unstandardized estimates, standardized parameter estimates ( $\beta$ ), and p-values for regression analyses of the relationship between the log of IL-8 and brain volumes/icv are presented. \*Denotes a result that was significant at p<0.05. †Denotes a result between p = 0.05 and p = 0.10.

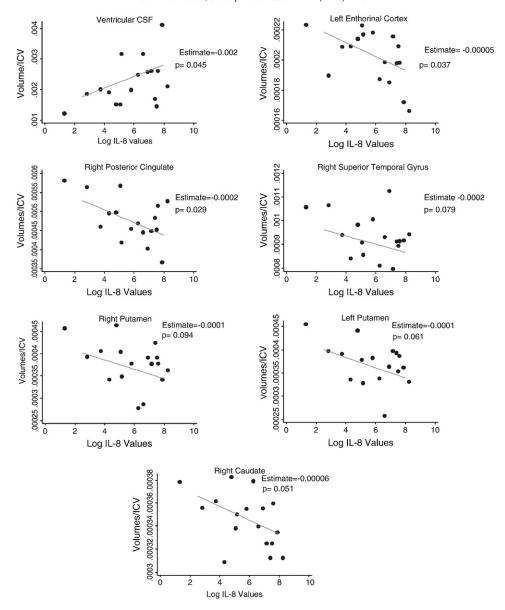


Fig. 3. Relationship between IL-8 and volumetric changes among schizophrenia cases controlling for maternal education. This figure displays raw ROI/icv values by log-transformed IL-8 values, as well as regression lines controlling for maternal education (maternal education was held constant at HS graduate or below for each regression line).

cance in the right superior temporal gyrus (p = 0.079), the right caudate (p = 0.051), and the putamen bilaterally (p = 0.061 and p = 0.094, right and left, respectively). There were no associations between fetal exposure to IL-8 and hippocampal volumes.

In secondary analyses among cases (Table 4 and Fig. 3), fetal exposure to increases in maternal IL-8 levels was related to volumetric decreases in the right posterior cingulate (p = 0.029). No significant relationships were observed between maternal IL-8 levels and any of the additional brain volumes examined among cases or in exploratory analyses among controls, although non-significant increases were observed for the right hippocampus (p = 0.061), right anterior cingulate (p = 0.097), and the right posterior cingulate (p = 0.088) among controls.

#### 4. Discussion

These results provide the first evidence that fetal exposure to increases in a maternal cytokine is associated with structural neuroanatomic alterations that have been consistently linked to schizophrenia (Wright et al., 2000). Among schizophrenia cases, increases in maternal IL-8 during the second/third trimesters of pregnancy were related to increases in ventricular CSF. In addition, we observed significant associations between maternal IL-8 and decreases in left entorhinal cortex and right posterior cingulate volumes, and volumetric decreases that approached significance in the right caudate, the putamen bilaterally, and the right superior temporal gyrus among cases.

 Table 4

 Results of unadjusted and adjusted regression analyses of maternal IL-8 and brain volumes in cases and controls: secondary analyses.

	Control participants (n = 8) unadjusted models				Control participants $(n=8)$ models adjusted for maternal education			
	Parameter estimate	95% CI	β	p-value	Parameter estimate	95% CI	β	p-value
Ventricular measures								
Sulcal cerebrospinal fluid	0.008	-0.012, 0.028	0.386	0.345	0.016	<b>−0.015, 0.046</b>	1.023	0.202
Cortical/limbic regions								
DLPFC (right)	0.0002	-0.0005, 0.0008	0.214	0.612	-0.0002	-0.002, 0.001	-0.324	0.674
DLPFC (left)	-0.0002	-0.0008, 0.0005	-0.232	0.580	-0.00007	-0.002, 0.002	-0.111	0.905
Inferior frontal cortex (right)	0.0003	-0.0005, 0.001	0.361	0.379	0.0002	-0.002, 0.002	0.221	0.806
Inferior frontal cortex (left)	0.0001	-0.0005, 0.0007	0.211	0.616	0.0003	-0.001, 0.002	0.582	0.512
Anterior cingulate (right)	0.0008	-0.0006, 0.002	0.500	0.207	0.0008	-0.0003, 0.002	0.779	0.097†
Anterior cingulate (left)	-0.002	-0.004, 0.0009	-0.527		-0.001	-0.007, 0.004	-0.423	0.547
Posterior cingulate (right)	0.0003	-0.0003, 0.001		0.252	0.0005	-0.0001, 0.001		0.088†
Posterior cingulate (left)	-0.0005	-0.001, 0.0004	-0.473		0.00005	-0.002, 0.002		0.947
Subcortical regions								
Thalamus (right)	0.0001	-0.0005, 0.0008	0.182	0.665	0.0006	-0.0008, 0.002	0.855	0.279
Thalamus (left)	0.0001	- 0.0004, 0.0006		0.602	0.0003	- 0.0006, 0.001		0.376
1471. it - /								
White/gray matter	0.000	0.040, 0.000	0.000	0.550	0.040	0.040.0047	0.005	0.005
White matter	0.006	-0.018, 0.030		0.578	-0.012	-0.042, 0.017	-0.625	
Cortical gray matter	-0.017	-0.041, 0.006	-0.594		-0.002	-0.046, 0.042	-0.077	
Subcortical gray matter	0.0002	-0.001, 0.001	0.163	0.700	0.0002	-0.002, 0.003	0.187	0.842
Brain region	Schizophrenia cases ( $n = 17$ ) unadjusted models				Schizophrenia cases education	(n=17) models adjus	ted for m	aternal
	Parameter estimate	95% CI	β	p-value	Parameter estimate	95% CI	β	p-value
Ventricular measures								
Sulcal cerebrospinal fluid	0.0004	<b>−</b> 0.006, 0.007	0.039	0.882	0.0006	− 0.005, 0.007	0.056	0.825
Cortical/limbic regions								
DLPFC (right)								0.010
DLITC (TIGHT)	-0.000004	-0.0004, 0.0004	-0.006	0.983	-0.00002	-0.0004, 0.0003	-0.025	
	-0.000004 $0.000004$			0.983 0.985	-0.00002 -0.00001		-0.025 $-0.015$	
DLPFC (left)	0.000004	-0.0004, 0.0004	0.005	0.985	-0.00001	-0.0004, 0.0004	-0.015	0.952
DLPFC (left) Inferior frontal cortex (right)	0.000004 0.0000004	-0.0004, 0.0004 -0.0003, 0.0003	0.005 0.001	0.985 0.998	-0.00001 $-0.00001$	-0.0004, 0.0004 -0.0003, 0.0003	-0.015 $-0.014$	0.952 0.956
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left)	0.000004 0.0000004 0.00001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003	0.005 0.001 - 0.026	0.985 0.998 0.922	- 0.00001 - 0.00001 - 0.00002	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0002	-0.015 $-0.014$ $-0.043$	0.952 0.956 0.866
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right)	0.000004 0.0000004 - 0.00001 - 0.0002	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0003 -0.0006, 0.00008	0.005 0.001 - 0.026 - 0.381	0.985 0.998 0.922 0.131	-0.00001 -0.00001 -0.00002 -0.0002	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007	-0.015 $-0.014$ $-0.043$ $-0.392$	0.952 0.956 0.866 0.123
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left)	0.000004 0.0000004 0.00001 0.0002 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.0004	0.005 0.001 - 0.026 - 0.381 - 0.148	0.985 0.998 0.922 0.131 0.571	-0.00001 -0.00001 -0.00002 -0.0002 -0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0002 - 0.0006, 0.00007 - 0.0007, 0.0004	-0.015 $-0.014$ $-0.043$ $-0.392$ $-0.157$	0.952 0.956 0.866 0.123 0.556
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (right)	0.000004 0.0000004 - 0.00001 - 0.0002	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.0004	0.005 0.001 - 0.026 - 0.381	0.985 0.998 0.922 0.131 0.571 0.026*	-0.00001 -0.00001 -0.00002 -0.0002	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007	-0.015 $-0.014$ $-0.043$ $-0.392$	0.952 0.956 0.866 0.123 0.556 0.029*
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (right) Posterior cingulate (left)	0.000004 0.0000004 0.00001 0.0002 0.0001 0.0002	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0003 -0.0006, 0.00008 -0.0006, 0.0004 -0.0003, -0.00002	0.005 $0.001$ $-0.026$ $-0.381$ $-0.148$ $-0.537$	0.985 0.998 0.922 0.131 0.571 0.026*	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0001 - 0.0002	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007 -0.0007, 0.0004 -0.0003, -0.00002	$-0.015 \\ -0.014 \\ -0.043 \\ -0.392 \\ -0.157 \\ -0.543$	0.952 0.956 0.866 0.123 0.556 0.029*
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (left) Posterior cingulate (left) Subcortical regions	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.0004 - 0.0003, - 0.00002 - 0.0003, 0.0001	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271	0.985 0.998 0.922 0.131 0.571 0.026* 0.292	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0001 - 0.0002 - 0.0001	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007 -0.0007, 0.0004 -0.0003, -0.00002 -0.0003, 0.0001	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (left) Posterior cingulate (left) Subcortical regions Thalamus (right)	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.00004 - 0.0003, - 0.00002 - 0.0003, 0.0001	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271	0.985 0.998 0.922 0.131 0.571 0.026* 0.292	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0001 - 0.0002 - 0.0001	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007 -0.0007, 0.0004 -0.0003, -0.00002 -0.0003, 0.0001	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (right) Posterior cingulate (left) Subcortical regions Thalamus (right)	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.0004 - 0.0003, - 0.00002 - 0.0003, 0.0001	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271	0.985 0.998 0.922 0.131 0.571 0.026* 0.292	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0001 - 0.0002 - 0.0001	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007 -0.0007, 0.0004 -0.0003, -0.00002 -0.0003, 0.0001	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (right) Posterior cingulate (left)  Subcortical regions Thalamus (right) Thalamus (left)  White/gray matter	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001 0.00001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.00004 - 0.0003, - 0.00002 - 0.0003, 0.0001 - 0.0001, 0.0001 - 0.00008, 0.0002	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271 0.048 0.193	0.985 0.998 0.922 0.131 0.571 0.026* 0.292	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0002 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0002 - 0.0006, 0.00007 - 0.0007, 0.0004 - 0.0003, - 0.00002 - 0.0003, 0.0001 - 0.0001, 0.0001 - 0.00009, 0.0002	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (left) Posterior cingulate (left) Subcortical regions	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.00004 - 0.0003, - 0.00002 - 0.0003, 0.0001	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271	0.985 0.998 0.922 0.131 0.571 0.026* 0.292	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0001 - 0.0002 - 0.0001	-0.0004, 0.0004 -0.0003, 0.0003 -0.0003, 0.0002 -0.0006, 0.00007 -0.0007, 0.0004 -0.0003, -0.00002 -0.0003, 0.0001	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266
DLPFC (left) Inferior frontal cortex (right) Inferior frontal cortex (left) Anterior cingulate (right) Anterior cingulate (left) Posterior cingulate (right) Posterior cingulate (left)  Subcortical regions Thalamus (right) Thalamus (left)  White/gray matter	0.000004 0.0000004 - 0.00001 - 0.0002 - 0.0001 - 0.0002 - 0.0001 0.00001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0003 - 0.0006, 0.00008 - 0.0006, 0.00004 - 0.0003, - 0.00002 - 0.0003, 0.0001 - 0.0001, 0.0001 - 0.00008, 0.0002	0.005 0.001 - 0.026 - 0.381 - 0.148 - 0.537 - 0.271 0.048 0.193	0.985 0.998 0.922 0.131 0.571 0.026* 0.292 0.854 0.459	- 0.00001 - 0.00001 - 0.00002 - 0.0002 - 0.0002 - 0.0002 - 0.0001	- 0.0004, 0.0004 - 0.0003, 0.0003 - 0.0003, 0.0002 - 0.0006, 0.00007 - 0.0007, 0.0004 - 0.0003, - 0.00002 - 0.0003, 0.0001 - 0.0001, 0.0001 - 0.00009, 0.0002	-0.015 -0.014 -0.043 -0.392 -0.157 -0.543 -0.285	0.952 0.956 0.866 0.123 0.556 0.029* 0.266 0.892 0.482

This table includes the results from the secondary multiple regression analyses with and without controlling for maternal education at birth. Unstandardized parameter estimates, 95% confidence intervals of the unstandardized estimates, standardized parameter estimates ( $\beta$ ), and p-values for regression analyses of the relationship between the log of IL-8 and brain volumes/icv are presented. \*Denotes a result that was significant at p<0.05. †Denotes a result between p = 0.05 and p = 0.10.

Ventricular enlargement is arguably the most well-replicated neuromorphologic anomaly in schizophrenia (Wright et al., 2000). Further, ventricular enlargement has been previously associated with hypoxia-associated OCs, suggesting that this structural change may have risk factors that are neurodevelopmental in origin (Cannon et al., 2002). Lastly, fetal exposure to activation of proinflammatory cytokines in rodents has been linked to increased ventricular CSF, which is the only known neuromorphologic volumetric

finding among animal studies to date (Patterson, 2009). Cumulatively, these findings suggest that fetal exposure to IL-8 may contribute to the known increases in ventricular volumes in schizophrenia.

Many of the other volumetric changes found in the present study also have been observed in schizophrenia and prodromal populations (Borgwardt et al., 2007; Pantelis et al., 2003; Wright et al., 2000). Specifically, parahippocampal and superior temporal gyrus (STG) volume reductions have been repeatedly

found among patients with schizophrenia and these regions have been shown to be decreased in the available prodromal studies, as well (Pantelis et al., 2003; Wright et al., 2000). Similar to the present study, in first-episode and prodromal patients basal ganglia volumes were decreased (Corson et al., 1999; Pantelis et al., 2003).

Interestingly, no significant associations were observed among controls. This pattern of null findings may support the assertion that a genetic or environmental factor associated with schizophrenia is necessary for IL-8 to exert damaging influences on the fetal brain, although studies with larger samples are necessary to confirm this possibility.

IL-8 is a proinflammatory chemokine produced by multiple cells involved in mobilizing, activating and degranulating neutrophils (Janeway et al., 2005). Although IL-8 is integral in the initial immune response to infection, elevations in IL-8 also are sometimes associated with other maternal conditions during pregnancy that increase risk for schizophrenia, such as preeclampsia, obesity and anemia (Basso et al., 2005; Dalman et al., 1999; Fain, 2006; Insel et al., 2008; Laskowska et al., 2007; Schaefer et al., 2000). It is possible that additional obstetric insults may add to, interact with, or precede the effects of IL-8 to cause disruptions in fetal neuronal development among cases, which should be teased apart in future studies.

There were several limitations in the present study that should be noted. Given the multiple tests conducted in this study, it is possible that spurious findings arose from type I error. It appears unlikely, however, that our results are entirely due to chance, as they are consistent with many previous studies in clinical and preclinical studies of schizophrenia. Further, there is the possibility of type 2 error, given the modest sample sizes. It should considered, however, that the present study is the first of its kind with access to serological evidence of prenatal events and follow-up brain imaging data. Nevertheless, it will be essential to attempt to replicate the results of the present study with larger samples.

In addition, the sample was limited to slightly more than 25% of cases from the overall cohort, raising the potential for selection bias. However, cases who participated in the DIBS study and cases from the original cohort who were not DIBS subjects did not differ with regard to several demographic variables and IL-8 levels; therefore, it is unlikely that case ascertainment bias accounts for our findings. Bias also would be mitigated by the fact that the sample was derived from a population-based study, in contrast to many clinical imaging studies, which draw upon hospital or clinic-based samples.

Lastly, the use of antipsychotics may have influenced our results. Although this is a possibility, IL-8 values were not significantly related to current antipsychotic use, which significantly decreases the likelihood that medication use confounded our findings. Nonetheless, long-term medication use, which is difficult to assess retrospectively, could have influenced the observed brain volumes.

In conclusion, the present study is the first to suggest that brain anomalies previously associated with schizophrenia may be partially attributable to in utero insults, such as fetal exposure to elevated IL-8. These findings support our previous association between fetal exposure to IL-8 and risk of schizophrenia in this birth cohort (Brown et al., 2004). Future research is necessary to replicate this finding and to delineate whether fetal exposure to IL-8 interacts with risk

genes for schizophrenia. Our findings underscore the potential importance of prenatal contributions to schizophrenia, with implications for prevention, early intervention, and treatment strategies.

#### Role of funding source

This study was supported by research grants to Dr. Brown from the National Institute of Mental Health (R01MH060249, R01MH63264, and K02 MH065422-06) and a postdoctoral NIMH schizophrenia research fellowship to Dr. Ellman (5 T32 MH018870-20) and grants N01HD13334 and N01HD63258 from The Eunice Kennedy Shriver Institute of Child Health and Human Development. These funding sources had no role in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the paper for publication.

#### Contributors

Dr. Ellman conducted all of the statistical analyses, contributed substantially to the conceptualization of the paper, study design, data interpretation, and was the main writer of the manuscript. Dr. Deicken supervised all of the imaging acquisition and imaging analyses for the study and contributed to writing of the manuscript. Drs Vinogradov, Kremen, and Poole contributed to the conceptual design, to data collection, data interpretation, and manuscript writing. Mr. Kern and Dr. Tsai aided in data management and consultation pertaining to statistical analyses, interpretation of the data, and contributed to manuscript writing. Dr. Schaefer contributed to case/control ascertainment and manuscript writing. Dr. Brown contributed to all areas pertaining to study conceptualization, study design, participant ascertainment, interpretation of the findings, and manuscript writing.

#### Conflict of interest

The authors have no financial disclosures and/or conflicts to report.

#### Acknowledgments

This study was supported by research grants to Dr. Brown from the National Institute of Mental Health (R01MH060249, R01MH63264, and K02 MH065422-06) and a postdoctoral NIMH schizophrenia research fellowship to Dr. Ellman (5 T32 MH018870-20) and grants N01HD13334 and N01HD63258 from The Eunice Kennedy Shriver Institute of Child Health and Human Development. We also thank Theo van Erp, Ezra Susser, Michaeline Bresnahan, Barbara Cohn, Nashid Chaudhury, Aundrea Cook, Justin Penner, Nicole Stephenson, Dawn Schpak and the study teams of the CHDS, the PDS, and the DIBS for their contributions to data collection and preparation of this manuscript. The authors also wish to acknowledge Larry Kegeles, M.D. for his helpful comments.

#### References

Basso, B., Gimenez, F., Lopez, C., 2005. IL-1beta, IL-6 and IL-8 levels in gynecoobstetric infections. Infect. Dis. Obstet. Gynecol. 13 (4), 207–211.

Borgwardt, S.J., Riecher-Rossler, A., Dazzan, P., Chitnis, X., Aston, J., Drewe, M., et al., 2007. Regional gray matter volume abnormalities in the at risk mental state. Biol. Psychiatry 61 (10), 1148–1156.

Brown, A.S., Derkits, E.J., 2010. Prenatal infection and schizophrenia: a review of epidemiologic and translational studies. Am. J. Psychiatry 167 (3), 261–280.

Brown, A.S., Hooton, J., Schaefer, C.A., Zhang, H., Petkova, E., Babulas, V., et al., 2004. Elevated maternal interleukin-8 levels and risk of schizophrenia in adult offspring. Am. J. Psychiatry 161 (5), 889–895.

Cannon, T.D., Kaprio, J., Lonnqvist, J., Huttunen, M., Koskenvuo, M., 1998. The genetic epidemiology of schizophrenia in a Finnish twin cohort: a population-based modeling study. Arch. Gen. Psychiatry 55, 67–74.

Cannon, T.D., van Erp, T.G., Rosso, I.M., Huttunen, M., Lonnqvist, J., Pirkola, T., et al., 2002. Fetal hypoxia and structural brain abnormalities in schizophrenic patients, their siblings, and controls. Arch. Gen. Psychiatry 59 (1), 35–41.

Cannon, T.D., Yolken, R., Buka, S., Torrey, E.F., 2008. Decreased neurotrophic response to birth hypoxia in the etiology of schizophrenia. Biol. Psychiatry 64 (9), 797–802.

Collins, D.L., Zijdenbos, A.P., Kollokian, V., Sled, J.G., Kabani, N.J., Holmes, C.J., et al., 1998. Design and construction of a realistic digital brain phantom. IEEE Trans. Med. Imaging 17 (3), 463–468.

- Corson, P.W., Nopoulos, P., Andreasen, N.C., Heckel, D., Arndt, S., 1999. Caudate size in first-episode neuroleptic-naive schizophrenic patients measured using an artificial neural network. Biol. Psychiatry 46 (5), 712–720.
- Dalman, C., Allebeck, P., Cullberg, J., Grunewald, C., Koster, M., 1999. Obstetric complications and the risk of schizophrenia: a longitudinal study of a national birth cohort. Arch. Gen. Psychiatry 56 (3), 234–240.
- Deicken, R.F., Pegues, M., Amend, D., 1999. Reduced hippocampal Nacetylaspartate without volume loss in schizophrenia. Schizophr. Res. 37 (3), 217–223.
- Deicken, R.F., Eliaz, Y., Chosiad, L., Feiwell, R., Rogers, L., 2002. Magnetic resonance imaging of the thalamus in male patients with schizophrenia. Schizophr. Res. 58 (2–3), 135–144.
- Ellman, L.M., Yolken, R.H., Buka, S.L., Torrey, E.F., Cannon, T.D., 2009. Cognitive functioning prior to the onset of psychosis: the role of fetal exposure to serologically determined influenza infection. Biol. Psychiatry 65 (12), 1040–1047.
- Fain, J.N., 2006. Release of interleukins and other inflammatory cytokines by human adipose tissue is enhanced in obesity and primarily due to the nonfat cells. Vitam. Horm. 74, 443–477.
- Fein, G., Di Sclafani, V., Tanabe, J., Cardenas, V., Weiner, M.W., Jagust, W.J., et al., 2000. Hippocampal and cortical atrophy predict dementia in subcortical ischemic vascular disease. Neurology 55 (11), 1626–1635.
- Gur, R.E., Keshavan, M.S., Lawrie, S.M., 2007. Deconstructing psychosis with human brain imaging. Schizophr. Bull. 33 (4), 921–931.
- Insel, B.J., Schaefer, C.A., McKeague, I.W., Susser, E.S., Brown, A.S., 2008. Maternal iron deficiency and the risk of schizophrenia in offspring. Arch. Gen. Psychiatry 65 (10), 1136–1144.
- Janeway, C.A., Travers, P., Walport, M., Schlomchik, M.J., 2005. Immunobiology: The immune system in health and disease, 6th ed. Garland, New York & London.
- Job, D.E., Whalley, H.C., Johnstone, E.C., Lawrie, S.M., 2005. Grey matter changes over time in high risk subjects developing schizophrenia. Neuroimage 25 (4), 1023–1030.
- Kwan, R.K., Evans, A.C., Pike, G.B., 1999. MRI simulation-based evaluation of image-processing and classification methods. IEEE Trans. Med. Imaging 18 (11), 1085–1097.
- Laskowska, M., Laskowska, K., Leszczynska-Gorzelak, B., Oleszczuk, J., 2007. Comparative analysis of the maternal and umbilical interleukin-8 levels

- in normal pregnancies and in pregnancies complicated by preeclampsia with intrauterine normal growth and intrauterine growth retardation. J Matern Fetal Neonatal Med 20 (7), 527–532.
- Manji, H.K., Moore, G.J., Chen, G., 2000. Clinical and preclinical evidence for the neurotrophic effects of mood stabilizers: implications for the pathophysiology and treatment of manic-depressive illness. Biol. Psychiatry 48 (8), 740–754.
- Nurnberger Jr., J.I., Blehar, M.C., Kaufmann, C.A., York-Cooler, C., Simpson, S.G., Harkavy-Friedman, J., et al., 1994. Diagnostic interview for genetic studies. Rationale, unique features, and training. NIMH Genetics Initiative. Arch. Gen. Psychiatry 51 (11), 849–859 (discussion 863–844).
- Pantelis, C., Velakoulis, D., McGorry, P.D., Wood, S.J., Suckling, J., Phillips, L.J., et al., 2003. Neuroanatomical abnormalities before and after onset of psychosis: a cross-sectional and longitudinal MRI comparison. Lancet 361 (9354), 281–288.
- Patterson, P.H., 2009. Immune involvement in schizophrenia and autism: etiology, pathology and animal models. Behav. Brain Res. 204 (2), 313–321.
- Schaefer, C.A., Brown, A.S., Wyatt, R.J., Kline, J., Begg, M.D., Bresnahan, M.A., et al., 2000. Maternal prepregnant body mass and risk of schizophrenia in adult offspring. Schizophr. Bull. 26 (2), 275–286.
- Schlotz, W., Phillips, D.I., 2009. Fetal origins of mental health: evidence and mechanisms. Brain Behav. Immun. 23 (7), 905–916.
- Susser, E.S., Schaefer, C.A., Brown, A.S., Begg, M.D., Wyatt, R.J., 2000. The design of the prenatal determinants of schizophrenia study. Schizophr. Bull. 26 (2), 257–273.
- Talairach, J., Tournoux, P., 1988. Co-planar stereotaxic atlas of the brain. Thieme Medical Publishers, New York.
- Wright, I.C., Rabe-Hesketh, S., Woodruff, P.W., David, A.S., Murray, R.M., Bullmore, E.T., 2000. Meta-analysis of regional brain volumes in schizophrenia, Am. J. Psychiatry 157 (1), 16–25.